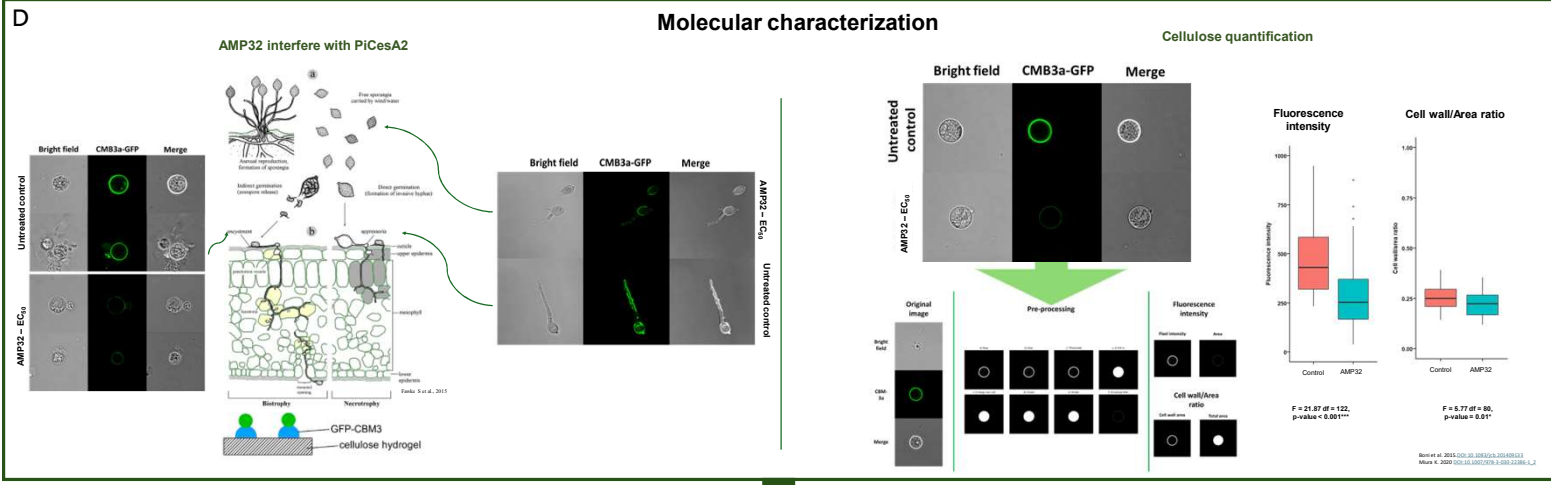
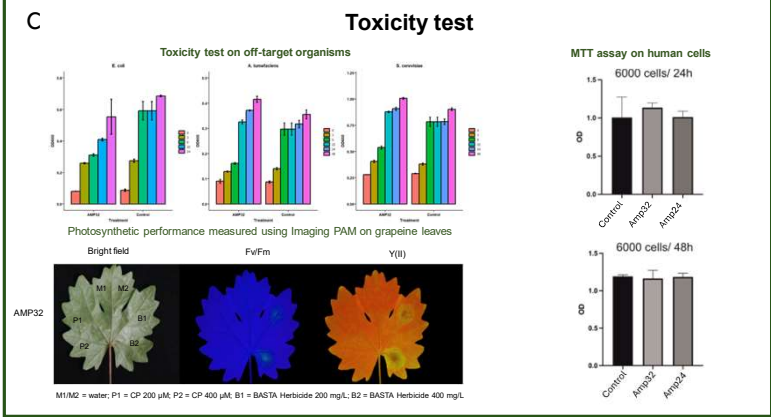
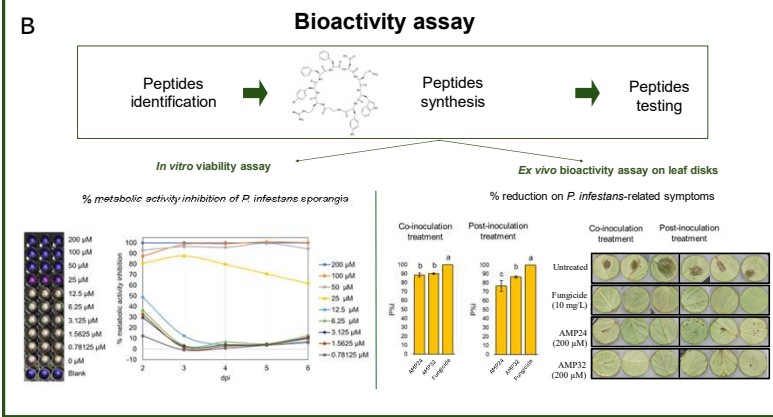
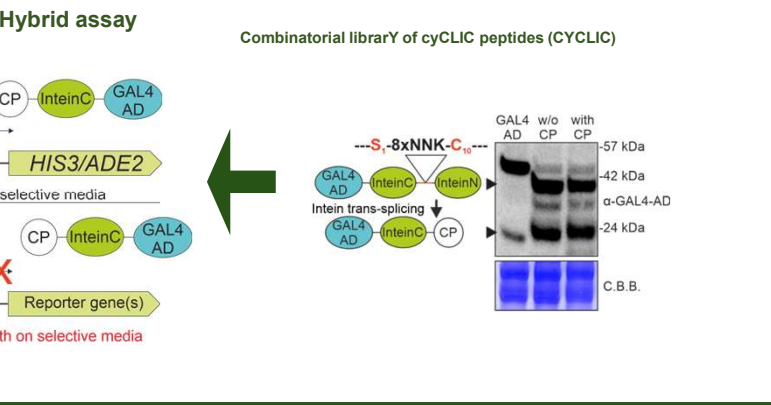
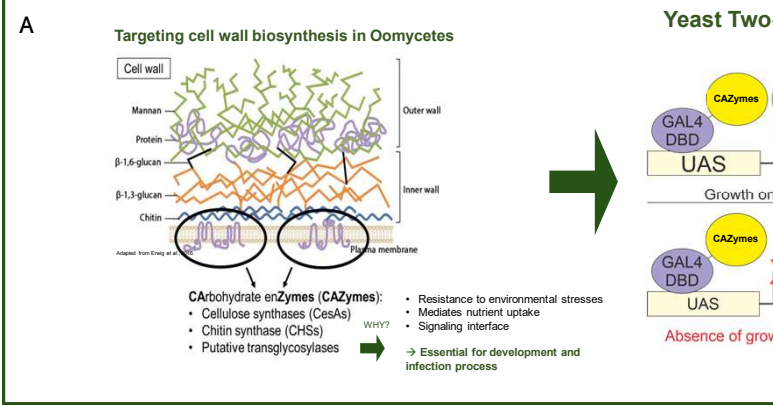
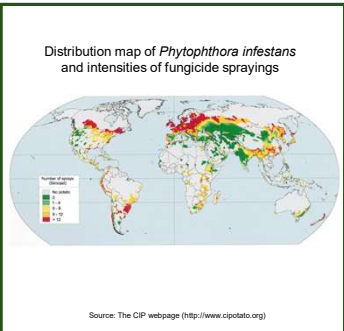


Abstract
Oomycetes, e.g. *Phytophthora infestans*, a widespread pathogen that causes late blight and affects most *Solanaceae* species, cause crop losses of several billion USD every year. Oomycetes life cycle consist in the production of sexual oospores and asexual zoospores, this event occurs numerous times during a single vegetative season. Defence against fungal pathogens is mainly based on the use of chemical fungicides¹, which give rise to great concerns for risks associated with human health, environmental toxicity, or fast development of resistance in target pathogens. Among those, several compounds have been banned or included in a list of candidates for substitution by the EU Commission, thus novel and sustainable molecules with low environmental impact are needed. Our group identified, through a Yeast Two-Hybrid-based combinatorial library of cyclic peptides (CYCLIC), an antimicrobial peptide (AMP32) able to bind a bait (i.e., target) of interest². This approach aims to increase the specificity toward the targeted protein and, as consequence, toward the desired pathogen, thus reducing the risks of negative effects on other organisms. In this work, the glycosyltransferase domain of the *P. infestans* Cellulose synthase 2 (PiCesA2) was selected as bait because of its essential role in cellulose biosynthesis and cell wall integrity³. Chemically synthesized AMP32 was tested for its antimicrobial activity in vitro and ex vivo assays and showed a strong *P. infestans* growth inhibition both in vitro and ex vivo. Noteworthy, the absence of toxicity in non-target organisms was also successfully addressed. Moreover, AMP32 showed similar antimicrobial effects when tested on other oomycete species with a high homology sequence for the PiCesA2, suggesting a broad-spectrum activity. A deeper investigation has confirmed that AMP32 compromises cell viability in *P. infestans*, showing a fungistatic activity and the ability to interfere with *P. infestans* cell membrane integrity. Other ongoing studies will deepen our understanding of the structural and biochemical insights of AMP32-PiCesA interaction and mode of action (MoA) and will provide useful information for peptide modification and chemical formulation suitable for field application.



Outcome: identification of peptides with antimicrobial activity for a sustainable agriculture

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