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## Background

*Pseudomonas aeruginosa* (*Pa*), a Gram-negative opportunistic bacteria, is the leading cause of nosocomial and chronic respiratory infections in people with cystic fibrosis (CF). *Pa* infections are frequently resistant to antibiotics. Phage therapy, leveraging the life cycle of lytic phages (viruses that kill bacteria), shows significant potential as a treatment for bacterial infections resistant to antibiotics.

A cocktail composed by four phages (CK4), able to kill *Pa* clinical strains isolated from people with CF, was recently developed. CK4 resistant mutants were isolated. All presented mutations in *wzy*, which is involved in the biosynthesis of the lipopolysaccharide (LPS) component of the outer membrane (OM)<sup>2</sup> (Fig. 1).

A CK4 component, i.e. the *Schitoviridae* DEV phage, can infect both the wt smooth strain producing long LPS with the O-antigen (Fig. 1, c), or rough strains producing short LPS variants with a truncated core (i.e. *algC*, *galU* and *wapH* mutants; tc in Fig. 1), whereas strains producing LPS containing a single O-antigen repeat (i.e. *wzy* mutants; u+1 in Fig. 1) are DEV resistant.

DEV gp53 gene encodes long tail fibers that are usually the receptor binding proteins of phages. A  $\Delta$ gp53 DEV mutant was constructed using CRISPR-Cas editing. The mutant phage can infect rough mutants, but cannot grow in PAO1 or *wzy* strains (Fig. 2)<sup>3</sup>.

## Hypothesis I

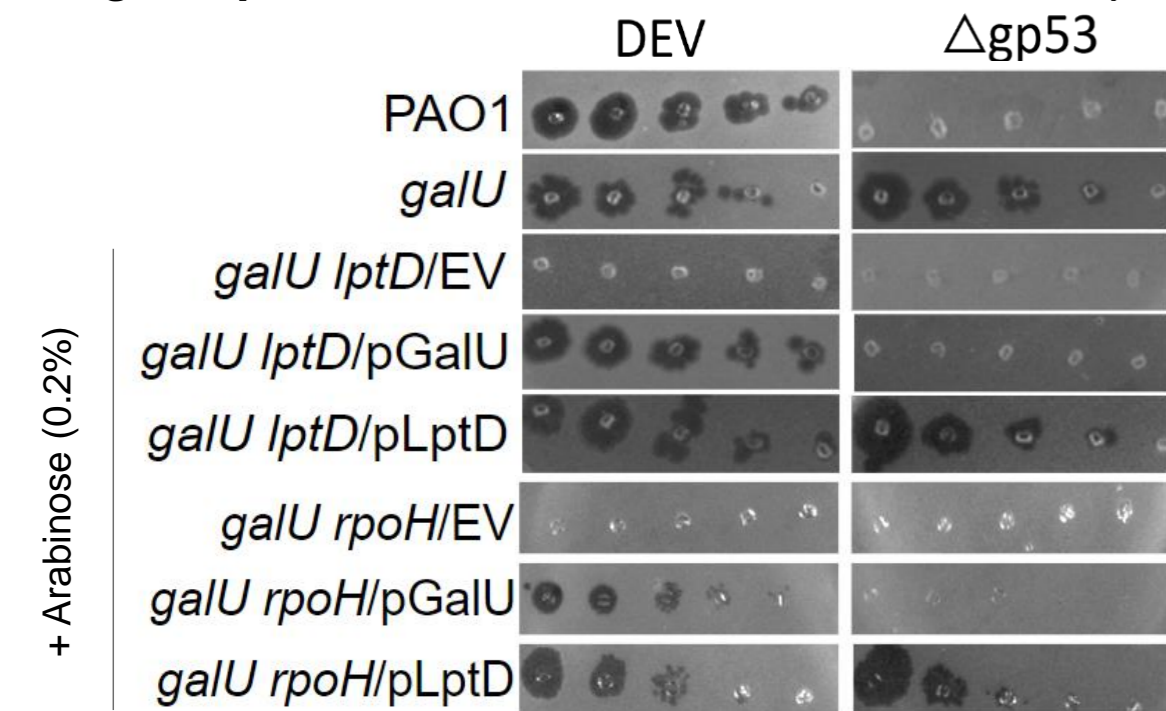
DEV may enjoy a two-receptor adsorption mechanism. The first receptor could be the O-antigen recognized by Gp53. The second receptor could be exposed in rough mutants, making the interaction with the O-antigen dispensable, and hidden, or absent, in *wzy* mutants.

## Results I

### 1. Isolation of DEV Resistant Double Mutants and complementation of the phage resistant phenotype.

Two spontaneous DEV resistant mutants of the phage-sensitive *galU* strain were isolated, carrying an extra mutation besides that in *galU*, namely:

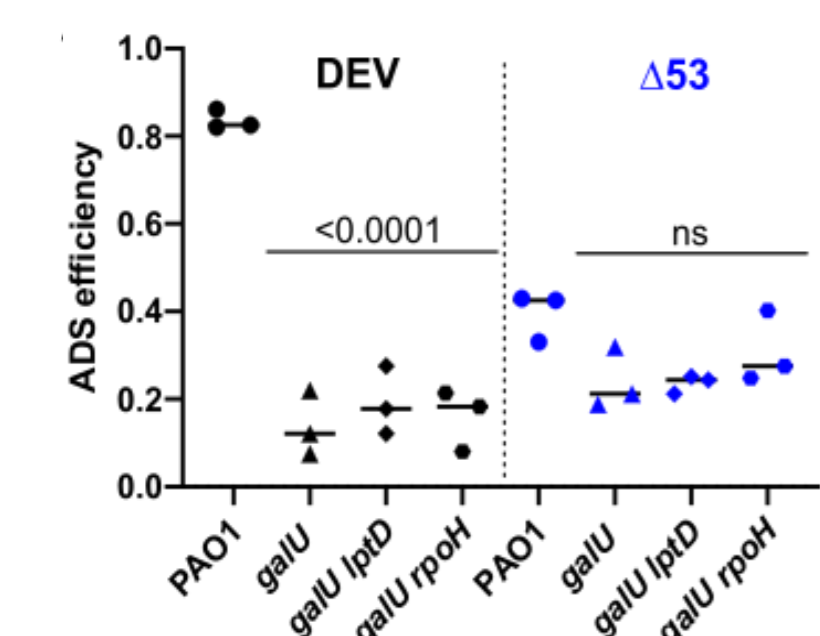
- ✓ ***galU lptD***: it contains a 33 bp long deletion in *lptD*, encoding the LPS OM transporter LptD.
- ✓ ***galU rpoH***: it contains a missense mutation in *rpoH* encoding  $\sigma^H$  sigma factor.



**Figure 3. Complementation assay of phage resistant mutants.** Our working hypothesis on adsorption: *galU lptD*/EV: Empty vector, *galU* and *lptD* mutations are present, no receptors available. *galU lptD*/pGalU: DEV uses the O-antigen as receptor.  $\Delta$ gp53 lacks tail fibers to attach to O-antigen. *galU lptD*/pLptD: DEV and  $\Delta$ gp53 use LptD as receptor.

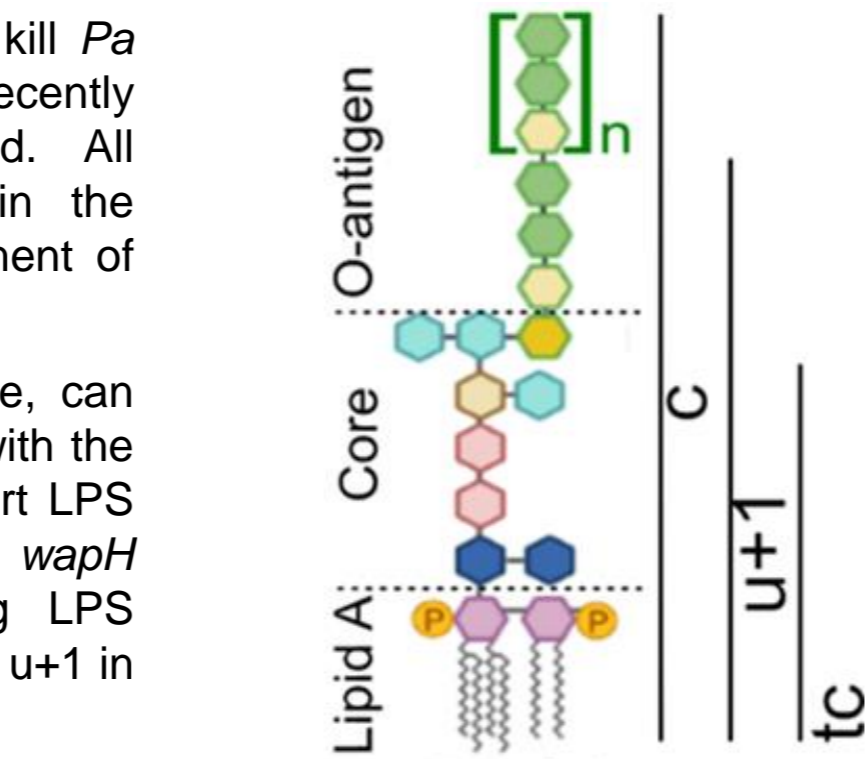
### 2. DEV and $\Delta$ gp53 Adsorption tests

To assess if adsorption is the defective step in resistant mutants

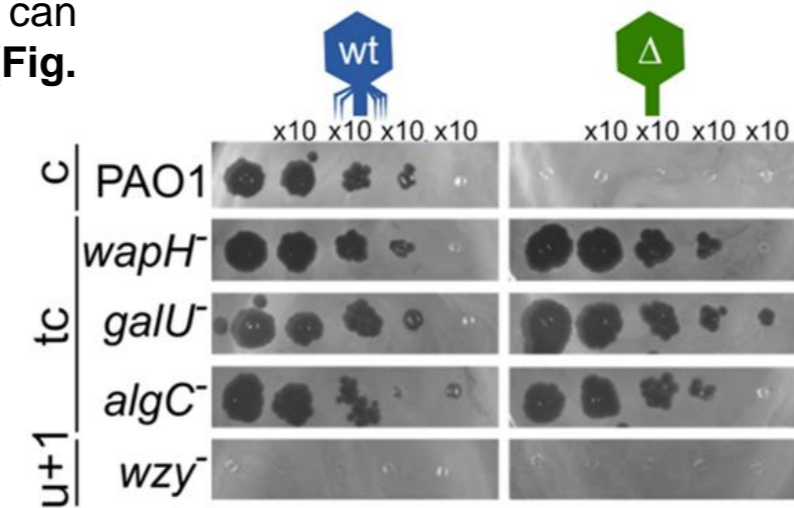


**Figure 4. Adsorption efficiency of DEV and  $\Delta$ gp53 to the indicated mutants (left) or to *galU lptD* double mutant carrying the indicated plasmids.** EV, empty vector, pGM931. The lines represents average. P calculated with One-way ANOVA and Šidák's multiple comparisons test is reported.

- The absence of O-antigen prevents stable adsorption by DEV.
- Consistently,  $\Delta$ gp53 always has inefficient adsorption increased when *lptD* is overexpressed.



**Figure 1. Predicted LPS structure in phage resistant mutants.**



**Figure 2. Replica Plating on *Pa* Mutants of DEV (wt) and  $\Delta$ gp53 ( $\Delta$ )**

In *galU lptD*:

- When *lptD* is complemented, adsorption of both phages is restored.
- When *galU* is complemented, only DEV adsorption is restored.

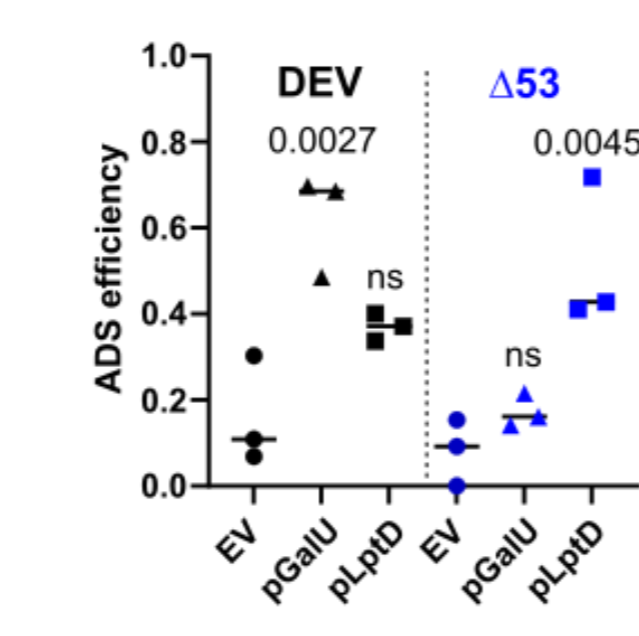
The *lptD* mutation does not prevent phage growth if a smooth LPS can be produced and the phage has Gp53 tail fibers.

In *rpoH lptD*:

- When *galU* is complemented, only DEV adsorption is restored.
- *lptD* overexpression suppresses the *rpoH* mutation.

The *rpoH* mutation interferes with adsorption, most likely by preventing the production or export to the OM of the second receptor.

To assess if adsorption is restored in presence of complementing plasmids



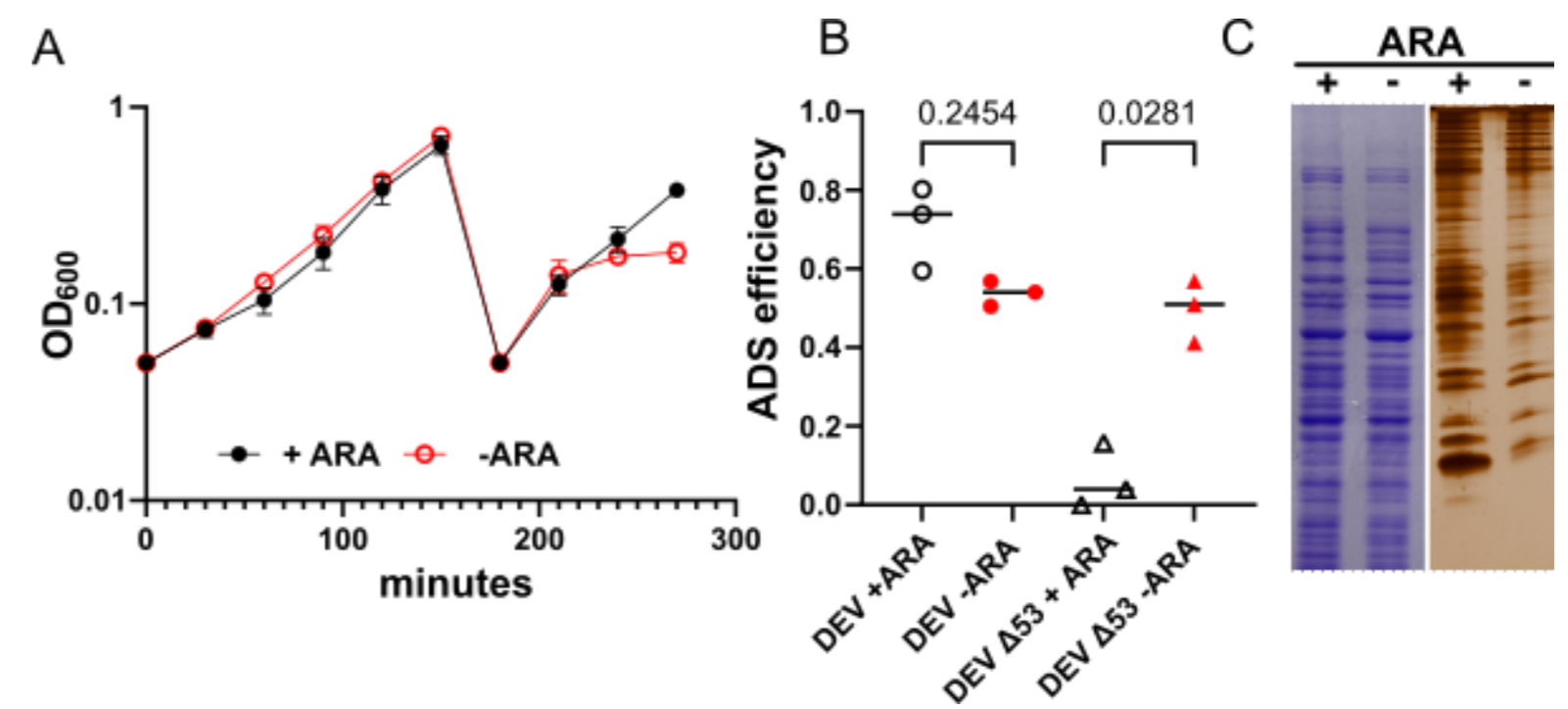
## Hypothesis II

The LptD protein is DEV secondary receptor. Alternatively, another LPS form not produced in the *lptD galU* mutant could be used by the phage as receptor.

## Results II

### 3. Adsorption to a *Pa* strain with LPS depletion

To evaluate if LPS is required for adsorption

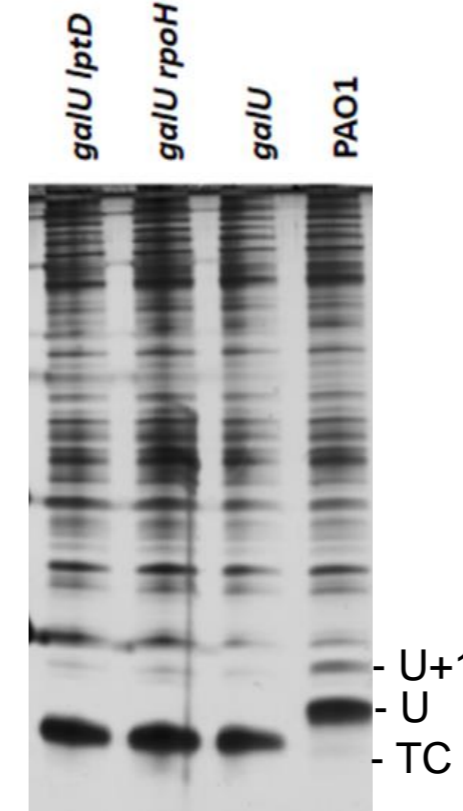


**Figure 5. LPS depletion stimulates DEV  $\Delta$ 53 adsorption.** A. Growth of the *araBp-lpxA* strain in presence or absence of 0.2% arabinose. The adsorption assay (B) and LPS analysis (C) were executed on bacteria collected at the last time point. B. Adsorption efficiency. Significance estimated with One-way ANOVA and Šidák's multiple comparisons test. C. LPS (right panel) and proteins (left panel).

- LpxA catalyzes one of the first steps of LPS biosynthesis.
- DEV can adsorb to LpxA-depleted cells in spite of severe LPS depletion.
- $\Delta$ p53 adsorption improved significantly upon LpxA (and LPS) depletion.

### 4. Analysis of LPS prepared from strains defective in LPS production or transport

To evaluate the effects of mutations on LPS pattern

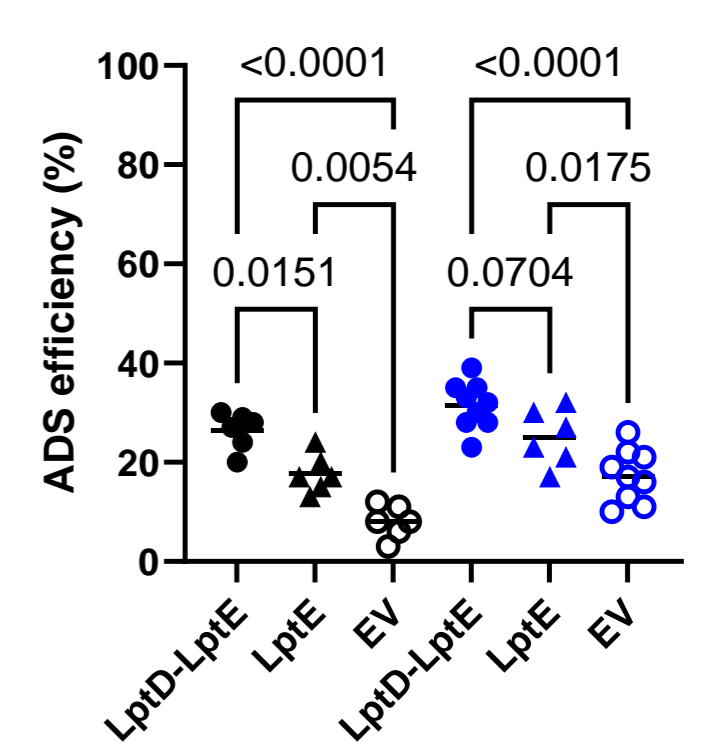


**Figure 5. LPS migration on 18% tricin gel and silver staining.**

*galU lptD* and *galU rpoH* double mutants show an LPS pattern almost identical to that of the *galU* single mutant.

### 5. Adsorption of DEV to a rough *E. coli* strain expressing *Pa lptD*

To assess whether LptD promote adsorption in a heterologous system



**Figure 6. Adsorption of DEV (black) and  $\Delta$ gp53 (blue) to *E. coli* JW3606 ( $\Delta$ *rfaG*) expressing the indicated *Pa* proteins.** EV, empty vector. Significance estimated with One-way ANOVA and Šidák's multiple comparisons test.

LptE forms a complex with LptD and is essential for the correct assembly of LptD on the OM. When both *Pa lptD* and *lptE* genes are expressed, the adsorption of DEV and  $\Delta$ gp53 to *E. coli* slightly but consistently increases.

## Conclusions

- DEV can adsorb either to the O-antigen with the Gp53 long tail fibers or to the secondary receptor with another unidentified receptor binding protein.
- DEV  $\Delta$ gp53 cannot bind the O-antigen. However it can bind the secondary receptor when not hindered by LPS capped with a single or multiple O-antigen repeats.
- Adsorption to the secondary receptor only is hardly detectable in conventional adsorption tests suggesting that it can be unstable.
- Data supports the essential LptD-LptE complex as the secondary receptor of DEV phage.

## Bibliography

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2. F. Forti, C. Bertoli, M. Cafora, S. Gilardi, A. Pistocchi and F. Briani. (2023) Identification and impact on *Pseudomonas aeruginosa* virulence of mutations conferring resistance to a phage cocktail for phage therapy. *Microbiology Spectrum* 12;11(6):e0147723.
3. Cingolani G, Lokareddy R, Hou CF, Forti F, Iglesias S, Li F, Pavlenok M, Niederweis M, Briani F. Integrative structural analysis of *Pseudomonas* phage DEV reveals a genome ejection motor. *Res Sq* [Preprint]. 2024. doi: 10.21203/rs.3.rs-3941185/v1.